# Stability and Behavior in Carbonate Cores for New Enhanced-Oil-Recovery Polymers at Elevated Temperatures in Hard Saline Brines

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### Summary

The goal of this work was to identify viable polymers for use in the polymer flooding of high-temperature carbonate reservoirs with hard, saline brines. This study extensively examined recent enhanced-oil-recovery (EOR) polymers for stability, including new 2-acrylamido-tertbutylsulfonic acid (ATBS) polymers with a high degree of polymerization, scleroglucan, *n*-vinylpyrrolidone (NVP)-based polymers, and hydrophobic associative polymers. For each polymer, stability experiments were performed over a 2-year period under oxygen-free conditions (less than 1 ppb) at various temperatures up to 180°C in brines with total dissolved solids (TDS) ranging from 0.69 to 24.4%, including divalent cations from 0.034 to 2.16%. Use of the Arrhenius analysis was a novel feature of this work. This rarely used method allows a relatively rapid assessment of the long-term stability of EOR polymers. Rather than wait years or decades for results from conventional stability studies at the reservoir temperature, reliable estimates of the time-temperature stability relations were obtained within 2 years. Arrhenius analysis was used to project polymer-viscosity half-lives at the target reservoir temperature of 99°C. The analysis suggests that a set of ATBS polymers will exhibit a viscosity half-life over 5 years at 120°C and over 50 years at 99°C, representing a novel finding of this work and a major advance for extending polymer flooding to higher temperatures.

For five polymers that showed potential for application at higher temperatures, corefloods were performed under anaerobic conditions. Another novel feature of this work was that anaerobic floods were performed without using chemical oxygen scavengers, chemical stabilizing packages, or chelating agents (that are feared to alter rock properties). Using carbonate cores and representative conditions, corefloods were performed to evaluate polymer retention, rheology in porous media, susceptibility to mechanical degradation, and the residual resistance factor for each of the polymers at 99°C.

#### Introduction

A significant fraction of the oil that could be recovered by polymer flooding and other chemical-flooding methods exists in hot carbonate reservoirs (Masalmeh et al. 2019). When common concentrations of divalent cations are present in a brine (5 to 10% of the total salinity), hydrolyzed polyacrylamide (HPAM) polymers eventually hydrolyze and precipitate in hot reservoirs, compromising their utility at greater than 70°C (Davison and Mentzer 1982; Zaitoun and Potie 1983; Moradi-Araghi and Doe 1987; Ryles 1988). If no dissolved oxygen or divalent cations are present, HPAM solutions can exhibit a viscosity half-life of 8 years at 100°C and 2 years at 120°C (Seright et al. 2010). If large amounts of soft water are available, HPAM solutions could (in concept) be effective in reservoirs up to 100°C if the polymer bank is sufficiently viscous to sweep through the reservoir with minimum mixing with the hard formation water (Maitin 1992; Seright et al. 2010). This concept requires control of ion exchange and carbonate dissolution so that too much hardness is not added to the polymer bank by interaction with formation minerals (Pope et al. 1978; Lake 1989; ten Berge et al. 2018). ten Berge et al. (2018) extensively examined ion exchange in the presence of HPAM, demonstrating that the polymer promotes dissolution of divalent cations.

However, in cases where use of soft water is impractical, more stable polymers are needed for applications in reservoirs that are hotter than 70°C. Since the 1980s, more stable synthetic polymers have been made by incorporating NVP and 2-acrylamido-2-methylpropanesulfonic acid (AMPS) or ATBS into acrylamide-based polymers (Doe et al. 1987; Moradi-Araghi et al. 1987). Until recently, issues with cost effectiveness limited their application (because of higher monomer cost and lower degree of polymerization than HPAM).

Within the past few years, polymer manufactures have proposed and evaluated several new or improved polymers for EOR use at elevated temperatures (Reichenbach-Klinke et al. 2011, 2013, 2018; Gaillard et al. 2010, 2014, 2015; Vermolen et al. 2011; Leonhardt et al. 2014; Siggel et al. 2014; Leblanc et al. 2015; AlZahid et al. 2016; Divers et al. 2017; Dupuis et al. 2017; Jensen et al. 2018; Rodriguez et al. 2018a, 2018b). Some recent work with these polymers has been reviewed by Alfazazi et al. (2018) and Delamaide (2018). Our paper examines the most promising polymers for potential use in high-temperature/salinity reservoirs with large spacing. Specifically, the envisioned application is a carbonate reservoir at 99°C, with formation salinity of 24.4% TDS. Less saline waters could be available for injection (i.e., "injection" water with 6.9%-TDS salinity and "smart" water with 0.69%-TDS salinity), but all have significant concentrations of divalent cations. The application relies on well spacings that are 1 to 2 km. With 1-km spacing, the interwell-polymer-transit time is expected to be approximately 17 years. For a polymer- or chemical-flooding process to be effective, the polymer must be sufficiently stable for a significant fraction of the expected interwell-transit time. To clarify, a 17-year transit time is approximated by dividing the average aqueous-phase pore volume (PV) in a pattern at residual oil saturation by the average injection rate. Of course, if the injected polymer channels inefficiently through the reservoir (e.g., through fractures), the actual interwell-transit time might be less. However, a prudent design would mitigate severe channeling before proceeding with the polymer flood.

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The largest well spacing for existing field applications of polymer flooding is approximately 300 m (Wassmuth et al. 2007; Wang et al. 2008, 2009). So, one might question the viability of using a well spacing of 1 to 2 km. Reduced well spacing could significantly speed up the response to the polymer flood and reduce the stability requirement for the polymer. However, for the target application, the operator has proprietary technical and economic reasons for insisting on large spacings.

Previous studies of polymer stability at elevated temperatures were commonly performed at the temperature of the target reservoir (Shupe 1981; Yang and Treiber 1985; Jabbar et al. 2017, 2018). Unfortunately, very few of these studies continued for more than 2 years. If the anticipated interwell-transit time is large compared with the life of the stability study, these studies cannot be expected to make credible predictions of long-term polymer effectiveness. To overcome this limitation, Arrhenius analyses can be used. By performing stability studies at a number of elevated temperatures, the Arrhenius analysis can establish the relation between viscosity, time, and temperature. Then, the Arrhenius equation can project polymer stability (e.g., viscosity half-life) at the target reservoir temperature. This approach was successfully used in two previous studies: one with xanthan (Seright and Henrici 1990) and one with HPAM with no divalent cations present (Seright et al. 2010). The primary criticism of the Arrhenius method is that predictions can be compromised if the mechanism of polymer degradation changes over the temperature range of the study (i.e., between the reservoir temperature and the maximum temperature of the study). However, Swiecinski et al. (2016), Sandengen et al. (2017, 2018), and Nurmi et al. (2018) recently examined the Arrhenius analysis for acrylamide-ATBS-based polymers and suggested that the method appeared valid.

This work will first provide a literature review of EOR-polymer stability at elevated temperatures. The goals of this literature review are to introduce the reader to previous stability studies of EOR polymers in the absence of dissolved oxygen; to note that most previous studies occurred only at the temperature of the intended application and occurred for a short time (2 years or less); and to provide an understanding of why most previous polymers (especially acrylamide-based polymers) were not sufficiently stable for high-temperature EOR applications. After the literature review, we detail the experimental methodology and the application of the Arrhenius analysis in assessing the stability of EOR polymers. Although the Arrhenius analysis is not new to reaction kinetics, the petroleum literature has very few reports directed at EOR-polymer stability (Seright and Henrici 1990; Seright et al. 2010). Without the Arrhenius analysis, reliable studies of polymer stability require many years or decades to perform.

In our research, the Arrhenius analysis assumes that the mechanism of polymer degradation remains basically unchanged over the temperature range of the study. Mechanisms envisioned for degradation for synthetic polymers include oxidative degradation; hydrolysis of side groups, followed by precipitation with divalent cations; crosslinking/gelation associated with side groups reacting with multivalent cations; and breakage of the polymer backbone by some other mechanism. For biopolymers, the envisioned mechanisms are much the same, although hydrolysis of side groups is unlikely to cause significant viscosity loss, while hydrolysis of the polysaccharide backbone is important (Seright and Henrici 1990). Because our studies were conducted in the complete absence of dissolved oxygen, oxidative degradation is eliminated as a mechanism. For polymers that have a moderate or high acrylamide or acrylate composition, the literature indicates that the dominant degradation mechanism (in the absence of oxygen) is the hydrolysis of side groups, followed by precipitation with divalent cations. Our stability studies will confirm this finding. For synthetic polymers that have low (or zero) acrylamide/acrylate content, we hope/speculate that the mechanisms of hydrolysis/precipitation/gelation will be minimized, leaving thermal breaking of the polymer backbone as the single/dominant mechanism for viscosity loss. With fewer mechanisms available for degradation, the Arrhenius analysis becomes more viable for long-term projections. In our experimental work, these ideas will be examined during long-term stability tests of the most promising EOR polymers. Stability tests were performed at four elevated temperatures (120, 140, 160, and 180°C) and then the Arrhenius analysis established the relation between viscosity, time, and temperature. Ultimately, the Arrhenius equation will project polymer stability (e.g., viscosity half-life) at the target reservoir temperature of 99°C. In contrast to studies with xanthan (Seright and Henrici 1990) and HPAM with no divalent cations present (Seright et al. 2010), the current study examines relatively new polymers in the presence of significant levels of divalent cations.

For five polymers that showed potential for application at higher temperatures, in addition to stability studies, this work also performed corefloods under anaerobic conditions. A procedure was developed to perform these floods with continuously monitored dissolved-oxygen concentrations, typically less than 5 ppb. This was performed without using chemical oxygen scavengers, chemical stabilizing packages, or chelating agents (that are feared to alter rock properties). Using representative carbonate cores and conditions, corefloods were performed to evaluate polymer retention, rheology in porous media, susceptibility to mechanical degradation, and the residual resistance factor at 99°C.

#### Literature Review of EOR-Polymer Stability at Elevated Temperatures

Degradation Mechanisms for Synthetic Polymers. At elevated temperatures, acrylamide-based polymers experienced degradation by at least three different mechanisms: amide/side-group hydrolysis, followed by precipitation with divalent cations; reaction with dissolved oxygen (resulting in cleavage of the carbon/carbon backbone); and breakage of the carbon/carbon backbone by some mechanism that does not involve dissolved oxygen. The literature on oxidative degradation of EOR polymers has been extensively reviewed elsewhere (Seright and Skjevrak 2015; Jouenne et al. 2017). Achieving less than 20 ppb dissolved oxygen is straightforward for most field applications where subsurface waters are used for the polymer makeup (Seright and Skjevrak 2015). Prudent engineering of surface equipment and leak prevention allows for the maintenance of near-zero dissolved-oxygen levels in the flow stream (Seright and Skjevrak 2015). For the low levels of dissolved oxygen that remain, contact with the reducing environment of a reservoir will quickly remove all dissolved oxygen (although some polymer degradation can be expected in hot reservoirs before the oxygen is depleted) (Seright et al. 2010). Polymer-stability results described in Seright and Henrici (1990) and Seright et al. (2010) were dominantly conducted without using chemical oxygen scavengers and "stabilization packages" (i.e., only nonchemical methods were used to remove oxygen), and yet (as described in those papers) they reported the greatest stability for xanthan and HPAM of any study to date. As noted by Seright and Skjevrak (2015), chemical oxygen scavengers and "stabilization packages" are only of value if significant dissolved oxygen is present. Seright et al. (2010) noted that even if ambient levels of dissolved oxygen are present (3 to 8 ppm), the highly reducing conditions and iron minerals (e.g., pyrite, siderite) in an oil reservoir will usually consume that oxygen within hours or days, even at low temperature (Xu et al. 2000). Because our studies were performed in the absence of dissolved oxygen, this topic will not be discussed further.

**Side-Group Hydrolysis.** HPAM polymers are known to be unstable at elevated temperatures if divalent cations are present (Davison and Mentzer 1982; Zaitoun and Potie 1983; Moradi-Araghi and Doe 1987; Ryles 1988). For temperatures greater than  $60^{\circ}$ C, even at neutral pH, acrylamide groups within the HPAM polymer experience hydrolysis to form acrylate groups. If significant concentrations of divalent cations (especially Ca<sup>2+</sup>) are present, HPAM polymers can precipitate if the fraction of acrylate groups (i.e., the degree of hydrolysis) in the polymer becomes too high. These facts limit the utility of HPAM polymers for many potential EOR applications in

warmer reservoirs. Moradi-Araghi and Doe (1987) suggested hardness limits in brines for various temperatures: 2,000 mg/L for  $75^{\circ}$ C, 500 mg/L for  $88^{\circ}$ C, and 270 mg/L for  $96^{\circ}$ C. For brines containing less than 20 mg/L of divalent cations, they suggest that polymer hydrolysis and precipitation will not be a problem for temperatures of  $204^{\circ}$ C or greater.

For the work in our paper, three brines were used: connate water, injection water, and smartwater. Connate water contained 180.5 g/L of sodium chloride (NaCl), 69.99 g/L of calcium chloride dihydrate (CaCl<sub>2</sub>·2H<sub>2</sub>O), and 21.4 g/L of magnesium chloride hexahydrate (MgCl<sub>2</sub>·6H<sub>2</sub>O) (243.571-g/L TDS). Injection water contained 58.296 g/L of NaCl, 10.56 g/L of CaCl<sub>2</sub>·2H<sub>2</sub>O, and 4.48 g/L of MgCl<sub>2</sub>·6H<sub>2</sub>O (68.975-g/L TDS). Smartwater contained 5.5782 g/L of NaCl, 10.564 g/L of CaCl<sub>2</sub>·2H<sub>2</sub>O, 0.4116 g/L of MgCl<sub>2</sub>·6H<sub>2</sub>O, 0.2617 g/L of sodium sulfate (Na<sub>2</sub>SO<sub>4</sub>), and 0.05099 g/L of sodium bicarbonate (NaHCO<sub>3</sub>) (6.898-g/L TDS). All brines were filtered through 0.45-µm filters before use. **Table 1** summarizes the water compositions used in this work.

lon	Smartwater	Injection Water	Connate Water
Na	2.293	22.928	70.991
Ca	0.288	2.880	19.080
Mg	0.054	0.536	2.561
$SO_4^{2-}$	0.177	0	0
$HCO_3^-$	0.037	0	0
TDS	6.898	68.975	243.571

Table 1—Water compositions (in g/L).

The hardness limit, from Moradi-Araghi and Doe (1987), of 270 mg/L for 96°C suggests that even the smartwater has too much calcium (288 mg/L) to avoid the hydrolysis/precipitation problem for normal HPAM polymers at 99°C. The limit proposed by Moradi-Araghi and Doe (1987) assumed that one-half the divalent-ion content was calcium and the other one-half was magnesium. Calcium is known to be much more active than magnesium in precipitating polymers (Moradi-Araghi and Doe 1987). Consequently, significant HPAM losses can be expected because of hydrolysis/precipitation at 99°C in all brines studied in this work.

Consideration has been given to the rate of amide hydrolysis in HPAM polymers. Data from Moradi-Araghi and Doe (1987) indicate that in 5% NaCl solution at 90 to 100°C, 60% degree of hydrolysis will be reached after 20 to 30 days. Depending on the temperature and divalent-ion concentration, HPAM precipitation can be anticipated by the time the 60% degree of hydrolysis is reached.

Examination of the literature indicates that the rate of amide hydrolysis is accelerated by the presence of neighboring acrylate groups (Swiecinski et al. 2016; Sandengen et al. 2017, 2018). However, for a given pH, similar kinetics are followed up to approximately 55% degree of hydrolysis. That suggests that for practical field applications, HPAM will probably end up with the same degree of hydrolysis deep in a 99°C reservoir, regardless of the HPAM chosen (i.e.,  $M_w$ , copolymerized, post-hydrolyzed, initial acrylamide/acrylate sequence, and so forth). Certain additives can affect the cloud point (temperature when HPAM becomes turbid from the phase transition), but these additives will probably be diluted or stripped before the polymer reaches very far into the reservoir. Thus, any commercially available HPAM is expected to experience hydrolysis and precipitation in the three brines for our current work. The most obvious method to make HPAM viable in these brines at 99°C is to reduce the divalent-cation content (especially calcium) to less than 130 mg/L.

Polymer precipitation can be overcome by copolymerizing acrylamide with more-stable monomers (such as AMPS/ATBS or NVP) that resist hydrolysis (Doe et al. 1987; Moradi-Araghi et al. 1987; Gaillard et al. 2010, 2014, 2015; Rodriguez et al. 2018a, 2018b; Divers et al. 2017; Dupuis et al. 2017; Zhang et al. 2017). These polymers have significantly improved resistance to precipitation; however, they are noticeably more expensive and less efficient viscosifiers than HPAM. Incidentally, our understanding is that AMPS and ATBS are synonyms for the same molecule.

Sandengen et al. (2017) studied the rates of acrylamide and ATBS hydrolysis for different acrylamide/ATBS copolymers. For initial ATBS content from 15 to 35% (i.e., 65 to 85% acrylamide), the number of acrylamide groups in the polymers decreased to 30% after 140 days at 100°C and to 5% after 60 days at 120°C (with the acrylamide being converted to acrylate). The half-life for ATBS hydrolysis was approximately 20 days at 120°C (when the initial ATBS content was 15 to 35%). ATBS hydrolysis was not apparent at 80°C, so the question is whether ATBS hydrolysis will be significant at 100°C for a practical EOR time frame (e.g., 5 years). Our interpretation/ extrapolation from this work is that the hydrolysis/precipitation problem (at 99°C in our three brines) cannot be avoided for ATBS (AMPS) contents lower than 40 to 50% (in the polymer).

Even if ATBS is used, there is uncertainty whether ATBS will be hydrolyzed significantly over 5 years at 99°C (Parker and Lezzi 1993). Sandengen et al. (2018) studied the stability of copolymers with up to 80% ATBS at temperatures of 120 and 140°C. For a polymer with 70% ATBS in synthetic seawater, the viscosity half-life was approximately 50 days at 120°C and 5.9 times faster at 140°C. If this trend is extrapolated to 100°C, the 70% ATBS polymer is projected to have a viscosity half-life of approximately 300 days. Sandengen et al. (2018) found that a polymer with 80% ATBS exhibited a viscosity half-life at 140°C that was four times longer than for a polymer with 70% ATBS. Extrapolating this trend suggests that a polymer requires at least 80% ATBS to provide a viscosity half-life longer than 3 years at 100°C. Of course, one can hope that this projection is pessimistic for 100°C. But note the comment of Sandengen et al. (2018): ATBS- and acrylamide-hydrolysis reactions "do not reach some equilibrium condition with remaining AM or ATBS." Thus, acrylamide/ATBS copolymers are expected to eventually hydrolyze to acrylate extensively at elevated temperatures.

A particularly useful observation from Sandengen et al. (2017) was that Arrhenius analysis (judging polymer stability from tests performed at various temperatures greater than the target temperature) was valid for HPAM-type polymers. Sandengen et al. (2018) also suggested that the rate of hydrolysis was not sensitive to total salinity.

A detailed examination was performed of several previous studies of stability of polymers for high-temperature EOR applications, including those from Doe et al. (1987), Vermolen et al. (2011), Gaillard et al. (2014, 2015), Zhuoyan et al. (2015), Dupuis et al. (2017), and Rodriguez et al. (2018b). This examination revealed that most previous studies occurred at only the temperature of the intended application, occurred for a short time (2 years or less), and were consistent with the points made previously: that high concentrations of stable monomers must be incorporated to improve the stability of synthetic EOR polymers.

**Degradation Mechanisms for Polysaccharides.** Polysaccharides (xanthan, scleroglucan, diutan, schizophyllan) are known to experience degradation by oxidation and hydrolysis. A literature review of these mechanisms (focused on xanthan degradation) was provided

by Seright and Henrici (1990). They noted that in the absence of dissolved oxygen, xanthan solutions exhibit their maximum stability at pH values between 7 and 8. At pH values less than 7, viscosity-decay constants decrease significantly with decreasing pH, indicating that acid-catalyzed hydrolysis might have an important role in xanthan degradation. Xanthan stability also drops sharply at pH values greater than 8 as solutions become more alkaline, suggesting that base-catalyzed fragmentation reactions might also be important. Their analysis indicated that under ideal conditions (no dissolved oxygen, pH 7 to 8, moderate to high salinities), a xanthan solution could maintain at least one-half of its original viscosity for a period of 5 years if the temperature does not exceed 75 to 80°C. Thus, xanthan was not deemed sufficiently stable for application at 99°C.

Some studies of stability of other EOR polysaccharides (scleroglucan, diutan, schizophyllan) suggest that these polymers have potential for application at higher temperatures. Jensen et al. (2018) reported insignificant viscosity loss for a scleroglucan biopolymer (in seawater) after 1 year of storage at 95°C. Also, in synthetic seawater, Davidson and Mentzer (1982) and Kalpakci et al. (1990) reported good stability for scleroglucan. In particular, Kalpakci et al. (1990) reported little or no viscosity loss for scleroglucan solutions over 720 days at 100°C. In contrast, Ryles (1988) noted that scleroglucan degraded within 3 months at 90°C. However, the stability of these polymers has not been examined using the Arrhenius analysis. Biodegradation is beyond the scope of this study.

## **Experimental Procedures**

**Long-Term-Stability Studies.** *Polymers.* During this work, 22 polymer candidates were examined from 14 companies. However, this paper will focus on the polymers that exhibited the greatest potential for stability at elevated temperatures. All polymers discussed in this paper are commercially available. **Table 2** provides a listing of the polymers examined. The first column lists our noncommercial code for a given polymer. The second column lists the polymer concentration (in ppm) to achieve approximately 25 cp (at 25°C and 7.3 seconds<sup>-1</sup>) in injection water (6.9% TDS with 0.288% calcium and 0.0536% magnesium). The third column provides what is publicly known regarding the composition of these polymers, while the fourth column indicates that all polymers were provided in powder form. Gaillard et al. (2014), Dupuis et al. (2017), and Rodriguez et al. (2018b) provide good discussions of the relative cost effectiveness of polymers containing ATBS and NVP. In general, the relative cost for a given synthetic polymer in Table 2 increases linearly with increased polymer concentration. It will also increase with increased ATBS content, and especially with increased NVP content.

	Amount to Achieve ~25 cp in 6.9%-TDS		
Polymer Code	Water (mg/L)	Polymer Composition	Product Form
HPAM6	1600	30% acrylate, 70% acrylamide, 18 million g/mol	Powder
AAMPS1	2750	25% ATBS, 75% acrylamide, 10–12 million g/mol	Powder
ATBSA	4100	>90% ATBS	Powder
ATBSB	2900	>90% ATBS, higher $M_w$	Powder
ATBST	3500	>90% ATBS, thermoresponsive	Powder
ATBS8A	3500	~80% ATBS	Powder
ATBS8B	2700	~80% ATBS	Powder
AMNVPATBS1	4400	50–65% acrylamide, 20–25% ATBS, 5–20% NVP, 3–5 million g/mol	Powder
NVP1	9000	Large % of NVP	Powder
HYASA	8000	Large % of unspecified stable monomer, small % of hydrophobic associative monomer	Powder
HYASB	8000	Large % of unspecified stable monomer, small % of hydrophobic associative monomer	Powder
SG1	950	Scleroglucan	Powder

Table 2—Polymers examined.

*Stability Methodology.* The viscosity targeted for the studies was 20 to 30 cp at 7.3 seconds<sup>-1</sup> and 25°C. Much of our methodology for assessing polymer stability can be found in Seright and Henrici (1990), Seright et al. (2010), and Seright and Skjevrak (2015). Polymer samples were prepared and viscosities, pH values, and dissolved-oxygen levels were measured inside an anaerobic chamber (Thermo Forma Anaerobic System Model 1025, Thermo Fisher Scientific, Waltham, Massachusetts, USA). **Fig. 1** shows one of these chambers. This unit continuously circulated an anaerobic gas (10 to 15% hydrogen and 85 to 90% nitrogen) through a palladium catalyst and a desiccant. Any free oxygen was reacted with hydrogen to form water, which was removed by the desiccant. Oxygen measurements were made with either a colorimetric metric method [CHEMets® (CHEMetrics, Midland, Virginia, USA) with a limit of oxygen detection between 1 and 5 ppb in aqueous solution] or a very sensitive dissolved-oxygen meter [Fibox 4 Trace (PreSens Precision Sensing GmbH, Regensburg, Germany), which is sensitive to 1 ppb oxygen in the liquid phase]. Under most circumstances, measurements indicated 0.000% oxygen in our chamber gas. As an exception, the chamber gas could rise to 0.035% oxygen immediately after moving items into the anaerobic chamber from the transfer chamber. As such, when anything was brought in from outside the main chamber, the transfer chamber was purged twice with pure-nitrogen gas and once with our anaerobic gas, interspersed with evacuations to a 65-kPa vacuum. Within 45 minutes of making a transfer, the oxygen content in the main chamber returned to 0.000%. Polymer solutions that were prepared in the anaerobic chamber typically contained no dissolved oxygen (i.e., less than 1 ppb).

Brines were mixed and filtered through 0.45- $\mu$ m filters (MilliporeSigma, Burlington, Massachusetts, USA) outside the anaerobic chamber. Then the brine was moved into the chamber, and a pump was used to bubble anaerobic gas through the brine. Less than 1 hour was required to drive the dissolved-oxygen content to less than 1  $\mu$ g/L. A vortex was formed using a magnetic stirrer, and powder-form synthetic polymer was added in the traditional manner and then mixed overnight at low speed. Note that the powder-form polymers were stored inside the anaerobic chamber. Scleroglucan was mixed using a blender inside the anaerobic chamber. Polymer-solution viscosities were measured inside the anaerobic chamber at 7.3 seconds<sup>-1</sup> and room temperature using a Brookfield Model DV-E viscometer with a UL adapter (Brookfield Ametek, Middleboro, Massachusetts, USA). After preparation, 70 cm<sup>3</sup> of polymer solution were placed in a 150-cm<sup>3</sup> stainless steel cylinder and sealed using stainless steel plugs with blemish-free threads that were

wrapped with yellow 3.5 mil gas-line nonstick tape. Normal white nonstick tape was inadequate. Then the cylinders were removed from the anaerobic chamber and placed in silicone oil baths (e.g., Thermo Neslab EX7, Thermo Electron Corporation, Newington, New Hampshire, USA) for various times at different temperatures ranging from 99 to 180°C. When a viscosity measurement was to be made, the cylinder was removed from the silicone bath, cooled rapidly in an ice bath, and then brought into the anaerobic chamber for viscosity, oxygen, and pH measurements at room temperature. After the measurements, the sample was returned to the same cylinder, resealed, removed from the anaerobic chamber, and returned to the appropriate silicone bath. An advantage of this method over previous flame-sealed glass-ampoule methods is that all measurements over time were made on the same polymer-solution sample. Also, pH and dissolved-oxygen measurements could readily be made on these samples. A disadvantage is that if the seal is compromised for the sample cylinders, the entire sample is lost. Fortunately, the technique has been perfected so that samples are rarely lost.



Fig. 1—Anaerobic chamber.

*Illustration of Stability Data Collected.* The Arrhenius method used in this work involves examining the stability of a given polymer (in a given brine) at four elevated temperatures (120, 140, 160, and 180°C) and then using the Arrhenius equation to project polymer stability at lower temperatures (especially at the target temperature of 99°C). To illustrate the data collected for each polymer, **Fig. 2** shows the stability to date for the polymer ATBSA at the four elevated temperatures in injection and connate water, respectively. (As mentioned previously, after a given time of storage at a given temperature, viscosity measurements were made at 25°C and a shear rate of 7.3 seconds<sup>-1</sup> for consistency, explaining the conditions listed on the *y*-axes of Fig. 2.) As of 621 days into the study, this polymer exhibits excellent stability in all three brines at 120°C. In all three brines, the viscosity half-life was roughly 1 year at 140°C, 50 days at 160°C, and approximately 8 days at 180°C. Achieving approximately 25 cp required 3100 mg/L in smartwater, 4100 mg/L in injection water, and 4200 mg/L in connate water. At temperatures of 140, 160, and 180°C, the studies have effectively reached completion because viscosities have decreased to low values. However, at 120°C, viscosities have not deteriorated much after 621 days. Thus, the data at 120°C must be projected to estimate the decay constant at that temperature.

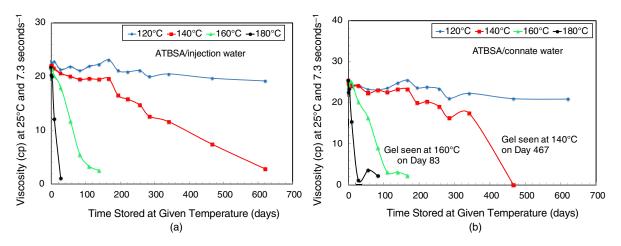


Fig. 2—Stability of ATBSA: (a) 4100 mg/L in injection water (6.9% TDS) and (b) 4200 mg/L in connate water (24.4% TDS).

In Fig. 2b, note that substantial gel was noted in the connate water after 83 days at  $160^{\circ}$ C and 467 days at  $140^{\circ}$ C. These observations suggest a change in the degradation mechanism at these times and temperatures. Presumably, before these indicated storage times, the polymer degraded by thermal breaking of the polymer backbone. However, at the indicated times, viscosity loss was accelerated by the onset of gel formation. Perhaps gel formation resulted after sufficient hydrolysis of side groups led to reaction with divalent cations in the very hard connate water. Apparently, this did not occur for the same polymer in the injection water (Fig. 2b). One could argue that the most appropriate data to use for the Arrhenius analysis are those acquired before the onset of the gelation mechanism (in the connate water) at a given temperature.

Arrhenius Projections of Polymer Stability at  $99^{\circ}C$ . The ultimate goal of this work is to estimate polymer stability at the reservoir temperature (99°C) using the experimental results obtained at higher temperatures (120, 140, 160, and 180°C). Use of the Arrhenius method allows for relatively rapid collection of stability data (at several temperatures hotter than the target reservoir) and projection of polymer stability for long time periods at lower temperatures (especially at the temperature of the target reservoir). Without this method, stability studies must be conducted for a substantial fraction of the life of the field project (many years). In the Arrhenius method, stability data vs. time (e.g., Fig. 2) are first fitted to

$$(\mu - \mu_s)/(\mu_o - \mu_s) = A_1 \exp(-t/\tau), \qquad (1)$$

where  $\mu$  is viscosity at time *t*,  $\mu_s$  is solvent viscosity, and  $\mu_o$  is original viscosity at time zero.  $A_1$  and  $\tau$  are fitting parameters. In particular,  $\tau$  is the decay constant (in days) for the degradation of a particular polymer solution. The  $\tau$  values can be related to temperature (*T*) through the Arrhenius equation,

 $1/\tau = A_2 \exp[-E/(RT)], \qquad (2)$ 

where *E* is the activation energy, *R* is the universal gas constant, and  $A_2$  is a pre-exponential parameter (a constant that is individual for a given reaction and sometimes indicative of the frequency of molecular interactions). Eq. 2 was used to fit the  $\tau$  values vs. temperature and to project  $\tau$  values at 99°C. The  $\tau$  values at 99°C were then converted to projected viscosity half-lives using

$$\text{Half-life} = \tau_{99^\circ\text{C}} \times \ln(2). \tag{3}$$

The right-most column of **Table 3** lists the projected half-lives for the various experiments, while the right-most column of **Table 4** lists the correlation coefficients from the fits to Eq. 2. For the case of ATBSA in injection water, **Fig. 3** provides a plot that illustrates how decay constants at 120 to  $180^{\circ}$ C are extrapolated to estimate the viscosity-decay constant at 99°C.

**Corefloods.** Five of the polymers from the stability studies were selected for examination during anaerobic corefloods at 99°C. Even though the three ATBS polymers (ATBSA, ATBSB, and ATBST from Table 2) appear most stable from the stability studies, polymers from different suppliers were selected to increase the diversity of the examination. All corefloods were performed at 100% water saturation.

*Polymers, Polymer Preparation, and Cores.* Five powder-form polymers examined in this work included: AMNVPATBS1 (as a benchmark), ATBSA, SG1 scleroglucan, NVP1, and HYASA. More detail for these polymers is given in Table 2 and during discussion of a given polymer.

Only injection brine was used in the corefloods. As mentioned previously, the brine was mixed and filtered through 0.45- $\mu$ m filters outside the anaerobic chamber. Then the brine was moved into the chamber (Fig. 1), and a pump was used to bubble anaerobic gas through the brine. Preparation of anaerobic polymer solutions was described previously. Because a greater amount of fluid transfer occurred during the corefloods, the level of dissolved oxygen present (2 to 22 ppb) was greater than during the long-term-stability studies (always less than 1  $\mu$ g/L).

**Table 5** lists the properties of the 3.8-cm-diameter carbonate cores used in this work. Polymer concentration and viscosity (cp at  $25^{\circ}$ C and 7.3 seconds<sup>-1</sup>) used in each core are also listed. Viscosities were measured using an MCR301 rheometer (Anton Paar GmbH, Graz, Austria) outside the anaerobic chamber and a digital viscometer (with UL adapter) inside the anaerobic chamber. (The viscosity listings in Table 5 used the MRC301 rheometer.)

*Coreflood Procedures.* Cores were saturated with injection brine (68.975 g/L of TDS), and porosity and initial permeability were measured under aerobic conditions. Then, cores were placed inside a biaxial Hassler core holder (Core Laboratories, Houston, Texas, USA) and pressurized to 1,300 psi (confining pressure) using anaerobic distilled water. Anaerobic fluids from inside the anaerobic chamber were piped directly into pumps (Teledyne ISCO, Lincoln, Nebraska, USA). From there, the fluids were forced through the core to confirm permeability and remove any dissolved oxygen from the core. Heat tape was used to heat and maintain the core and flow lines at 99°C. Effluent from the core was piped through an ice bath (to return the fluids to nearly room temperature) and then back into the anaerobic chamber. Anaerobic brine was flushed through the core and back into the anaerobic chamber until the dissolved-oxygen content of the effluent was less than 3 ppb. With this arrangement, anaerobic corefloods were performed without any chemical oxygen scavengers, chelators, or other chemicals that might change the character of the rock.

Next, for the polymer-retention test, the chosen anaerobic polymer solution was piped from the anaerobic chamber into a separate ISCO pump (that had previously been made anaerobic). This polymer solution contained 20 mg/L of potassium iodide as a tracer. The anaerobic polymer solution was then forced through the core at a rate of 2 ft/D (1 ft/D for the AMNVPATBS1 flood). Depending on the flood, between 11 and 21 PV of polymer solution were injected, while continuously monitoring the pressure drop across the core. A Retriever® 500 mini fraction collector (Teledyne ISCO, Lincoln, Nebraska, USA) was inside the anaerobic chamber to collect effluent samples in 3- to 4-cm<sup>3</sup> increments. Dissolved oxygen was regularly monitored during effluent collection. After collection, caps were placed on the sample tubes to prevent evaporation and to maintain oxygen-free conditions. After collection of all effluent-retention samples, they were removed from the anaerobic chamber for subsequent analysis. For a given sample tube, the cap was removed, the sample was weighed, viscosity was measured using a Vilastic Scientific (Austin, Texas, USA) VE rheometer (at 25°C and 7.3 seconds<sup>-1</sup>), and then the sample was placed back in the tube. After all the viscosities were measured, each sample was measured for potassium iodide content using a Thermo Scientific<sup>TM</sup> GENESYS 8 spectrophotometer at a wavelength of 230 nm. Finally, the samples were analyzed for total organic carbon (TOC) and nitrogen content using a TOC-L with TNM-L unit (Shimadzu Corporation, Kyoto, Japan) for nitrogen chemiluminescence. Nitrogen content is the most reliable means to detect synthetic (HPAM-type) polymers. TOC was used to detect polymer for SG1 scleroglucan (because the molecule does not contain nitrogen). Polymer-retention values were calculated as described in Manichand and Seright (2014) and Wang et al. (2020).

Polymer	Concentration (mg/L)	Water/TDS (%)	120°C τ (days)	140°C τ (days)	160°C τ (days)	180°C τ (days)	Projected Half-Life at 99°C (years)
ATBSA	3100	Smart/0.69%	2,595	266	40	5	61.1
ATBSA	4100	Injection/6.9%	2,691	266	50	5	54.8
ATBSA	4200	Connate/24.4%	1,947	400	37	9	67.3
ATBSB	2700	Smart/0.69%	3,942	350	30	6	63.1
ATBSB	2900	Injection/6.9%	3,990	462	16	7	133.3
ATBSB	3100	Connate/24.4%	4,359	200	18	1	126.9
ATBST	2800	Smart/0.69%	2,097	113	17	6	67.8
ATBST	3500	Injection/6.9%	2,583	292	20	8	57.6
ATBST	3800	Connate/24.4%	2,390	420	12	6	81.5
ATBS8A	2500	Smart/0.69%	306	49	12	3	4.1
ATBS8A	3500	Injection/6.9%	265	42	10	3	3.6
ATBS8A	3600	Connate/24.4%	287	55	11	5	4.5
ATBS8B	2200	Smart/0.69%	209	30	5	2	3.6
ATBS8B	2700	Injection/6.9%	124	15	5	3	1.4
ATBS8B	2900	Connate/24.4%	36	23	3	2	0.4
AMNVPATBS1	3500	Smart/0.69%	438	410	238	3	1.3
AMNVPATBS1	4400	Injection/6.9%	17.4	3.2	1.0	0.7	0.1
AMNVPATBS1	4500	Connate/24.4%	8.6	3.1	0.9	0.7	0.1
NVP1	7000	Smart/0.69%	864	633	368	186	3.5
NVP1	9000	Injection/6.9%	839	570	350	149	3.7
NVP1	9100	Connate/24.4%	689	197	68	31	5
HYASA	6000	Smart/0.69%	635	436	247	126	2.6
HYASA	8000	Injection/6.9%	633	371	236	23	2.2
HYASA	7000	Connate/24.4%	634	157	59	19	5.4
HYASB	6000	Smart/0.69%	519	432	258	57.5	3.3
HYASB	8000	Injection/6.9%	90.1	56.0	25	5	0.8
HYASB	6500	Connate/24.4%	21.0	4.6	2.2	0.8	0.3
SG1	920	Smart/0.69%	50	0.3	0.2	0.2	2
SG1	950	Injection/6.9%	20	2	0.2	0.2	1
SG1	880	Connate/24.4%	20	2	0.2	0.2	1

Table 3-  $\tau$  values (days from Eq. 1 fit) and 99°C half-life projections (years from Eq. 2 fit).

After completion of the retention test, the polymer solution was forced through the core at a series of decreasing rates to assess rheology in porous media and susceptibility to mechanical degradation. The highest rate was used first, with the intent of achieving roughly 1,000 psi across the core. This value was chosen so as not to exceed the 1,300-psi confining pressure. The flow rate and pressure drop were allowed to stabilize. Then, an effluent sample was collected within the anaerobic chamber for subsequent viscosity measurement (using the digital viscometer inside the chamber). Next, the injection rate was cut in half, and the process was repeated. This process was continued until the pressure drops were too low to measure reliably. The range of Darcy velocities extended from 139 to 0.017 ft/D for AMNVPATBS1 in 153-md rock, from 88 to 0.4 ft/D for ATBSA in 145-md rock, from 10.6 to 0.16 ft/D for ATBSA in 24-md rock, from 280 to 0.5 ft/D for SG1 scleroglucan in 106-md rock, from 528 to 1 ft/D for NVP1 in 322-md rock, and from 846 to 0.2 ft/D for HYASA in 504-md rock. The velocity ranges were determined by polymer rheology, core permeability, the 1,000-psi upper-pressure limit, and lower-pressure-limit accuracy for the transducers.

After completion of the rheology/mechanical-degradation study, 1.5 L (78 to 134 PV) of brine were injected and a residual resistance factor was measured.

## Results

**Long-Term Stability Studies.** The middle columns of Table 3 list  $\tau$  values (viscosity-decay constants in days, associated with Eq. 1) for various experiments (primarily those polymers with the greatest stability from this study), while Table 4 lists correlation coefficients from the fits to Eq. 1. In examining Tables 3 and 4, note that correlation coefficients are the lowest (worst) for cases where the polymer is still early in the study life (where the polymer solution has not lost much viscosity yet), such as the ATBS polymers at 120°C. Naturally, the correlation coefficients will rise and confidence in the 99°C projections will increase as a given study matures. An indication of the reproducibility of the results in Table 3 is implied by the consistency of the decay constants and projected viscosity half-lives in the three brines for the polymers: ATBSA, ATBSB, and ATBST. Because all three of these polymers have nearly the same composition (more than 90% ATBS), similar stability results were expected, as observed. Additional evidence of the reliability of projections is indicated by the correlation coefficients in Table 4.

		120°C	140°C	160°C	180°C	99°C Projection
Polymer	Water/TDS (%)	Eq. 1	Eq. 1	Eq. 1	Eq. 1	Eq. 2
ATBSA	Smart/0.69%	0.807	0.947	0.968	0.984	0.999
ATBSA	Injection/6.9%	0.836	0.928	0.988	0.980	0.998
ATBSA	Connate/24.4%	0.810	0.904	0.792	0.595	0.965
ATBSB	Smart/0.69%	0.585	0.833	0.959	0.959	0.977
ATBSB	Injection/6.9%	0.749	0.919	0.903	0.968	0.982
ATBSB	Connate/24.4%	0.712	0.476	0.888	0.981	0.964
ATBST	Smart/0.69%	0.769	0.806	0.894	0.862	0.996
ATBST	Injection/6.9%	0.744	0.906	0.904	0.979	0.988
ATBST	Connate/24.4%	0.671	0.854	0.846	0.857	0.965
ATBS8A	Smart/0.69%	0.987	0.989	0.987	0.997	0.999
ATBS8A	Injection/6.9%	0.982	0.978	0.910	0.998	0.999
ATBS8A	Connate/24.4%	0.972	0.990	0.843	1.000	0.999
ATBS8B	Smart/0.69%	0.976	0.979	0.997	0.999	0.996
ATBS8B	Injection/6.9%	0.942	0.971	0.979	0.633	0.985
ATBS8B	Connate/24.4%	0.727	0.797	0.790	1.000	0.939
AMNVPATBS1	Smart/0.69%	0.831	0.804	0.753	0.993	0.990
AMNVPATBS1	Injection/6.9%	0.976	0.993	0.726	0.922	0.996
AMNVPATBS1	Connate/24.4%	0.967	0.990	0.941	0.891	0.997
NVP1	Smart/0.69%	0.749	0.831	0.925	0.876	0.980
NVP1	Injection/6.9%	0.808	0.833	0.822	0.686	0.974
NVP1	Connate/24.4%	0.589	0.832	0.852	0.917	0.998
HYASA	Smart/0.69%	0.822	0.827	0.921	0.861	0.987
HYASA	Injection/6.9%	0.838	0.809	0.731	0.959	0.903
HYASA	Connate/24.4%	0.725	0.952	0.920	0.955	0.996
HYASB	Smart/0.69%	0.795	0.806	0.928	0.962	0.903
HYASB	Injection/6.9%	0.917	0.921	0.750	0.755	0.954
HYASB	Connate/24.4%	0.875	0.624	0.989	0.953	0.976
SG1	Smart/0.69%	0.977	1.000	1.000	1.000	0.825
SG1	Injection/6.9%	0.978	0.982	1.000	1.000	0.953
SG1	Connate/24.4%	0.953	0.911	1.000	1.000	0.932

Table 4—Correlation coefficients from fits to Eqs. 1 and 2.

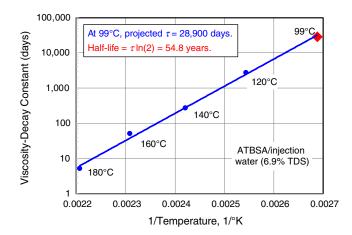


Fig. 3—Projection of decay constants at high temperatures to estimate stability at 99°C.

**Coreflood Results.** *Polymer Retention.* Fig. 4 shows the results of the polymer-retention studies for the six anaerobic corefloods at 99°C. The cores were initially saturated with only injection water (6.8975% TDS, no oil).

Polymer retention was assessed using the methodology described in Manichand and Seright (2014) and Wang et al. (2020). Basically, polymer retention derives from the difference in area between the breakout curves for tracer (blue curves in Fig. 4) and polymer (black, green, and red curves). In Fig. 4, the black curves are effluent carbon (from TOC analysis), the green curves (except for

scleroglucan) are effluent nitrogen (from chemiluminescent nitrogen), and the red curves are effluent viscosity (measured using the VE rheometer at  $25^{\circ}$ C and 7.3 seconds<sup>-1</sup>). For reasons described in Wang et al. (2020), inaccessible PV was zero, and the nitrogen curves were the most reliable indicator of effluent-polymer concentration. (As an exception, because scleroglucan does not contain nitrogen, effluent carbon was used instead. In Fig. 4d, the green curve indicates pressure drop across the core.) Polymer-retention values are given in Fig. 4 and also listed in **Table 6**. Calculated polymer-retention values were 211 µg/g for ATBSA in 145-md carbonate, 415 µg/g for ATBSA in 24-md carbonate, 204 µg/g for NVP1 in 322-md carbonate, 911 µg/g for AMNVPATBS1 in 153-md carbonate, 1126 µg/g for HYASA in 504-md carbonate, and more than 1250 µg/g for SG1 scleroglucan in 106-md carbonate. As will be discussed later, these are all considered high retention values.

Polymer	AMNVPATBS1	ATBSA	ATBSA	SG1	NVP1	HYASA
Core	272	28	15	264	24	324
Weight (g)	118	143.0	131.8	113.2	136.3	146.7
Length (cm)	5.35	6.66	6.20	5.15	6.57	6.08
$k_w$ (md)	153	145	24	106	322	504
$\phi$	0.187	0.258	0.281	0.257	0.26	0.28
Concentration (mg/L)	4400	4100	4100	950	9000	8000
Viscosity (cp) at 7.3 seconds <sup>-1</sup>	20.8	21.0	22.6	34.8	20.6	26.7

Table 5—Core properties and polymer concentrations for experiments.

*Plugging during Polymer Injection.* Except for the case with scleroglucan (Fig. 4d), pressures stabilized quickly (within 2 PV) during polymer injection. Thus, no progressive plugging was noted during most corefloods. In contrast, the green curve in Fig. 4d reveals that injection pressure rose continuously during injection of 14 PV of SG1 scleroglucan. After 14 PV, the effluent viscosity was only 6% of the injected value, while the effluent carbon was only 30% of the injected value. These results indicated very slow propagation of the SG1 polymer during the flood. As mentioned previously, the calculated retention value exceeded 1250  $\mu$ g/g. At the end of the experiment, a thick layer of slimy gel was noted on the injection face (**Fig. 5**), indicating that the polymer was stripped from the solution at the injection face. The polymer was dissolved very well before injection. There was no undissolved polymer (as was true for all six corefloods).

*Resistance Factors.* Resistance factor is defined as brine mobility (before polymer injection) divided by polymer-solution mobility. It is a measure of the effective viscosity of the polymer solution in porous media (relative to water). The seventh row in Table 6 lists the stabilized resistance factors during the retention experiments of Fig. 4. For comparison, the eighth row lists initial viscosity (at 25°C and 7.3 seconds<sup>-1</sup>). For the cases of AMNVPATBS1, ATBSA in 145-md rock, NVP1, and HYASA, resistance factors were roughly similar to the initial viscosity listing. This observation also suggests that little or no plugging (either on the face or internally within the core) occurred for these cases. For the case of ATBSA in 24-md rock, the resistance factor (56) was 2.5 times greater than the viscosity listing, suggesting that some level of plugging occurred. For the case of SG1 scleroglucan, no resistance factor was calculated because severe face plugging was noted.

*Viscosity Losses during Injection during Retention Experiments.* Near the end of each retention experiment in Fig. 4, we noted the fractional loss of viscosity upon propagation through the cores. As indicated in the ninth row of Table 6, these losses were modest or small (i.e., 1 to 9% loss) for AMNVPATBS1, ATBSA (in both permeabilities), and HYASA. For NVP1, the loss was 23%, which seems high considering that NVP polymers were thought to be very stable (Doe et al. 1987). However, our observations during the long-term stability study also noted relatively rapid loss of part of the NVP-solution viscosities at elevated temperatures. For SG1 scleroglucan, the effluent viscosity was only 6% of the injected value because of the stripping of the polymer from solution (as mentioned previously).

*Rheology and Mechanical Degradation in Porous Media.* After the retention test, the polymer solution was injected at a sequence of rates to examine mechanical degradation and rheology in porous media. **Figs. 6 and 7** show the results. At the highest rate (139 ft/D) for AMNVPATBS1 (black curves in Figs. 6 and 7), the resistance factor was 29.6 and the effluent viscosity was 33% less than the injected value (largely because of mechanical degradation). As the rate was decreased to 8.7 ft/D, the resistance factor decreased to 13.7. For rates of 34 ft/D and less, the effluent viscosity was consistently 9% less than the injected value (attributed to thermal degradation). As rates decreased to less than 8.6 ft/D, resistance factors increased (to 59.8 at 1 ft/D). During this part of the experiment, the resistance factor at 1 ft/D (59.8) was significantly greater than the value (28.4) observed during the retention experiment. This result suggests some plugging occurred over the course of the rheology/mechanical-degradation portion of the experiment. Overall, the black curves in Figs. 6 and 7 suggest that in 153-md rock, the onset of shear thickening occurred at approximately 9 ft/D, and the onset of mechanical degradation occurred between 35 and 70 ft/D.

At the highest rate (88 ft/D) for ATBSA in 145-md rock (solid red curves in Figs. 6 and 7), the resistance factor was 53.5 and the effluent viscosity was 5% less than the injected value. As rate was decreased to 11 ft/D, the resistance factor decreased to 17.4. For rates of 44 ft/D and less, no significant degradation was apparent. As rates decreased to less than 11 ft/D, resistance factors increased (to approximately 24 at 1 to 2 ft/D). During this part of the experiment, the resistance factor at 2 ft/D (approximately 24) was somewhat less than the value (34) observed during the retention experiment. No explanation is apparent for the difference. Overall, the red curves in Figs. 6 and 7 suggest that in 145-md rock, the onset of shear thickening occurred at approximately 11 ft/D, and the onset of mechanical degradation occurred between 44 and 88 ft/D.

At the highest rate (10.6 ft/D) for ATBSA in 24-md rock (red curves with red circles in Figs. 6 and 7), the resistance factor was 81.8 and the effluent viscosity was 14% less than the injected value. As the rate decreased to 1.3 ft/D, the resistance factor decreased to 55.9. For rates of 2.6 ft/D and less, no significant degradation was apparent. As rates decreased to less than 1.3 ft/D, the resistance factors were fairly constant. During this part of the experiment, the resistance factor at 2 ft/D (approximately 56) was roughly the same the value (59.6) observed during the retention experiment. Overall, the results suggest that in 24-md rock, the onset of a very modest shear thickening occurred at approximately 1.3 ft/D, and the onset of mechanical degradation occurred at approximately 5 ft/D.

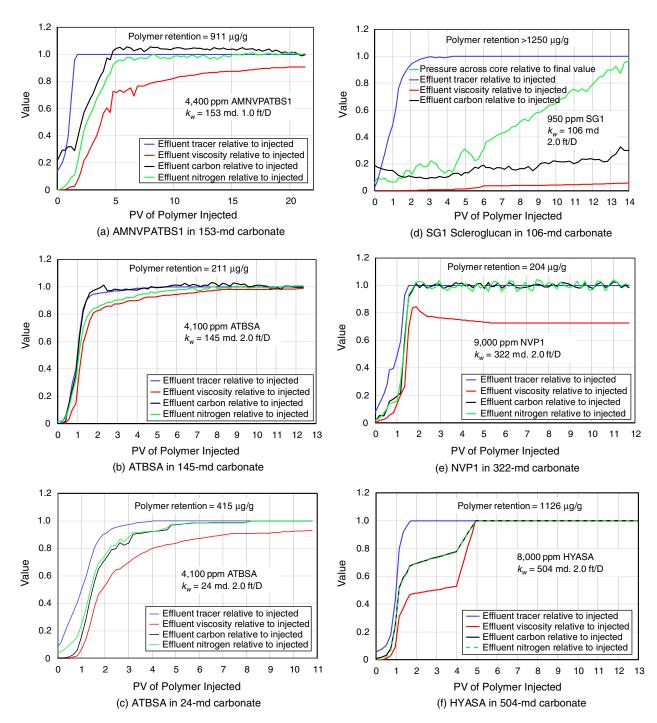


Fig. 4—Polymer-retention results at 99°C, initially saturated with injection water (6.8975% TDS) and under anaerobic conditions.

At the highest rate (280 ft/D) for SG1 scleroglucan (green curves in Figs. 6 and 7), the resistance factor was 15.1 and the effluent viscosity was the same as the injected value. As the rate was decreased to 70 ft/D, the resistance factor increased to 43.7. Over this range, the effluent viscosity was the same as the injected value, indicating no mechanical degradation and that the polymer could propagate through the core at these high rates. For lower rates, the apparent shear-thinning behavior continued, but the effluent viscosity decreased as rates were reduced to less than 18 ft/D, indicating that the polymer was filtered from the solution at the lower rates. The extremely high resistance factors at the lowest rates (3,300 at 0.55 ft/D) were attributable to face plugging, not the actual rheology in porous media. Thus, in agreement with previous work at low temperatures, SG1 scleroglucan shows shear-thinning rheology in porous media. At 99°C, the polymer propagates effectively and shows shear-thinning behavior in the carbonate cores at high rates. However, at moderate and low rates at 99°C, SG1 scleroglucan plugs the carbonate rock.

At the highest rate (528 ft/D) for NVP1 (blue curves in Figs. 6 and 7), the resistance factor was 14.9 and the effluent viscosity was the same as the injected value. Thus, no mechanical degradation was noted, even at this very high rate. As the rate was decreased to 66 ft/D, the resistance factor decreased to 3.7, indicating that the onset of a mild shear thickening occurred at approximately 66 ft/D. As rates decreased from 66 to 4 ft/D, resistance factors were fairly similar (averaging approximately 4), although an odd "hump" appears for the blue curve of Fig. 6 with a peak at 16 ft/D. This is attributable to a single data point and might not be a real effect. As rates were further reduced to 1 ft/D, the resistance factor increased to 17.3. At 9000 mg/L of polymer, NVP1 required noticeably higher concentrations than the other polymers in providing the viscosity and the resistance factor. The blue curve in Fig. 7 indicates increased viscosity loss as

injection rate decreased. This behavior occurred because slower rates lead to longer exposure time (to  $99^{\circ}$ C), and therefore greater thermal degradation at the slower rates. It was not caused by mechanical degradation (because there was no viscosity loss at the highest rates), and it was not caused by stripping of polymer (because the effluent-nitrogen and -carbon values were at the injected levels).

Polymer	AMNVPATBS1	ATBSA	ATBSA	SG1	NVP1	HYASA
Core	272	28	15	264	24	324
Permeability (md)	153	145	24	106	322	504
Concentration (mg/L)	4400	4100	4100	950	9000	8000
Retention (µg/g)	911	211	415	>1250	204	1126
Plugging observed while injecting?	No	No	No	Yes	No	No
Resistance factor at 1–2 ft/D	28.4*	34	56	Plugs	28.6	24.3
Initial viscosity (cp) at 7.3 seconds <sup>-1</sup>	20.8	21.0	22.6	34.8	20.6	26.7
Viscosity loss while injecting at 1–2 ft/D	9%	1%	7%	70%	23%	1%
Onset of mechanical degradation (ft/D)	35	44	5	_	_	_
Onset of shear thickening (ft/D)	9	11	1.3	_	66	53
Residual resistance factor	2.2	2.2	6.5	176	4	2.3

Table 6—Summary of anaerobic coreflooding results at 99°C in carbonate cores.



Fig. 5—SG1 gel on the injection face at the end of the coreflood.

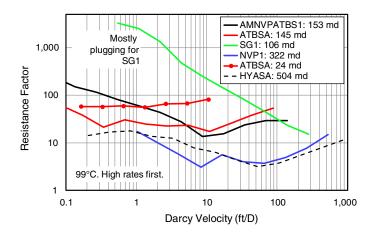


Fig. 6—Resistance factors vs. Darcy velocity for polymers in anaerobic carbonate cores at 99°C.

At the highest rate (846 ft/D) for HYASA (dashed black curves in Figs. 6 and 7), the resistance factor was 11.3 and the effluent viscosity was the same as the injected value. Thus, no mechanical degradation was noted, even at this very high rate. As the rate was decreased to 53 ft/D, the resistance factor decreased to 3.2, indicating that the onset of shear thickening occurred at approximately 53 ft/D in 504-md rock. As the rates decreased from 53 to 0.8 ft/D, the resistance factors were fairly similar (averaging approximately 4). As the rates were further reduced to 3.3 ft/D, the resistance factor increased to 12.6. For lower rates (down to 0.2 ft/D), the resistance factor remained fairly constant.

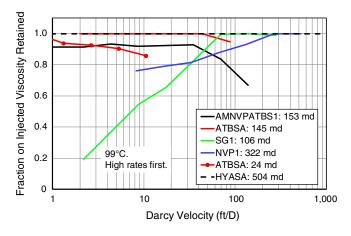


Fig. 7—Viscosity losses after injection for polymers in anaerobic carbonate cores at 99°C.

**Residual Resistance Factors.** The residual resistance factor is defined as the brine mobility before polymer divided by brine mobility after polymer has been displaced. It is a measure of the permeability reduction caused by the retention of a polymer. However, when no internal pressure taps are present (as in our experiments), any face plugging makes the residual resistance factors appear artificially high. In our work, the experiment with SG1 scleroglucan showed obvious signs of face plugging (Fig. 5). In the other corefloods, no polymer or gel was noted on the injection sand faces at the end of the experiments.

Previous work (Seright 2017; Wang et al. 2020) demonstrated that residual resistance factors can be artificially high if insufficient brine is flushed through the core. Specifically, more than 100 PV of brine might be required to displace all mobile polymer and achieve a true residual resistance factor.

For AMNVPATBS1, 134 PV of injection brine was flushed through the core, yielding a final residual resistance factor of 2.2. These results might suggest a small amount of plugging, which is consistent with the observations associated with the 1-ft/D points discussed previously for the black curve in Fig. 6. For ATBSA, 79 PV of brine flush yielded final residual resistance factors of 2.2 in 145-md rock and 6.5 in 24-md rock. With SG1 scleroglucan, 102 PV of brine flush resulted in a final residual resistance factor of 176, indicating a substantial amount of face plugging. After 79 PV of brine, the final residual resistance factor was 4.0 for NVP1. For HYASA, flushing with 78 PV of brine ended with a final residual resistance factor of 2.3.

### Discussion

**Long-Term Stability Discussion.** *HPAM (HPAM6).* Partially HPAMs are the most cost-effective polymer for low-salinity, low-temperature applications. Unfortunately, the literature indicates no hope that these polymers will be sufficiently stable at 99°C in the field connate or injection waters because they will hydrolyze and precipitate. Some hope was maintained that HPAM might be stable in the smartwater, although the criteria from Moradi-Araghi and Doe (1987) indicated that the brine must not contain more than 135 mg/L of calcium and 135 mg/L of magnesium at our target temperature. (Smartwater contains 288 mg/L of calcium.) Consequently, tests were performed with HPAM6 (a common commercial HPAM with a  $M_w$  of 18 million g/mol and 30% degree of hydrolysis) (1600 mg/L, 25 cp at 7.3 seconds<sup>-1</sup> and 25°C) to confirm. Results confirmed that the HPAM precipitated quickly at elevated temperatures (less than 3 days at 120°C), even in smartwater. The precipitate was seen as a white layer at the bottom of the storage container. TOC analysis (Shimadzu TOC-VCSH) confirmed that more than 80% of the polymer precipitated from the solution. Total nitrogen analysis (Shimadzu TOC-VSCH) revealed that the ammonia that emanated from the HPAM remained free in the solution. These results confirm that no HPAM will be sufficiently stable in any of the three field brines at elevated temperatures. The only way to make HPAM sufficiently stable is to remove divalent cations (especially calcium) from the waters to achieve a calcium concentration less than 135 mg/L [from Moradi-Araghi and Doe (1987)].

Acrylamide/ATBS Copolymer (AAMPS1). To reduce the hydrolysis/precipitation problem mentioned previously, copolymers of acrylamide and ATBS (or AMPS) were proposed. An acrylamide/ATBS copolymer, AAMPS1, with  $M_w$  of 6 to 8 million g/mol and 25% AMPS was obtained. For such chemistry, the ATBS monomer is slightly more expensive than acrylamide, and incorporation of ATBS does not allow achieving  $M_w$  values as high as for HPAM. Consequently, ATBS copolymers usually require more polymer to achieve a given viscosity. The stability of AAMPS1 was studied in the three field brines at temperatures ranging from 120 to 180°C. Initial solutions were clear. White precipitates were noted after a few days in all brines stored at temperatures of 140°C or higher. A white precipitate was also noted in the connate water after 6 days at 120°C. Significant viscosity losses occurred for all brines at all temperatures. TOC analysis confirmed loss of polymer from solution. Sandengen et al. (2017) and Moradi-Araghi and Doe (1987) project that the hydrolysis and precipitation problem will eventually occur at 99°C for acrylamide/ATBS polymers with less than 40 to 50% ATBS, which is the case for the brines under consideration here. Because AAMPS1 contains only 25% ATBS, it does not appear that this polymer will have sufficient stability for use in high-temperature/salinity and large-spacing applications. Because of (a speculated) concern of a major change in the degradation mechanism between 100 and 120°C, the stability of AAMPS1 was examined specifically at 99°C. The red curves in **Fig. 8** illustrate the poor stability of AAMPS1 in the two most saline field brines at 99°C. At Day 50, it was already apparent that AAMPS1 gelled and was not sufficiently stable at 99°C in connate water.

*Scleroglucan (SG1).* Scleroglucan is a neutral extracellular polysaccharide produced by the fungus *Athelia (Sclerotium) rolfsii.* The polysaccharide shares the backbone chemical structure of polymerized glucose. Its chemical structure consists of a linear  $\beta(1-3)$  D-glucose backbone, with one  $\beta(1-6)$  D-glucose side chain for every three main residue. This polymer shares much in common with schizophyllan. Scleroglucan is suggested to have a triple-helix structure in solution, thus hopefully shielding the polymers from chemical attack and increasing stability over other polysaccharides (Fournier et al. 2018).

In the three brines (smart, injection, connate), fits of the Arrhenius equation to the 120 and  $140^{\circ}$ C scleroglucan-viscosity-loss data projected a viscosity half-life of approximately 1 year at 99°C. The projection fits well with the actual behavior observed in the injection water at 99°C (blue curve in Fig. 8a). The polymer is interesting in that only 880 to 950 mg/L of polymer are needed in the three

brines to achieve the target viscosity (approximately 25 cp). During our stability experiments at the highest temperatures (160 to  $180^{\circ}$ C), some of this scleroglucan biopolymer appeared to be "cooked" to a black residue. It is possible that this cooked material was cellular debris (associated with the fermentation process) that was not actually scleroglucan. Other than a small loss caused by the cooking of the polymer, the TOC analysis indicated that the polymer stayed in the solution, regardless of the salinity or hardness of the brine.

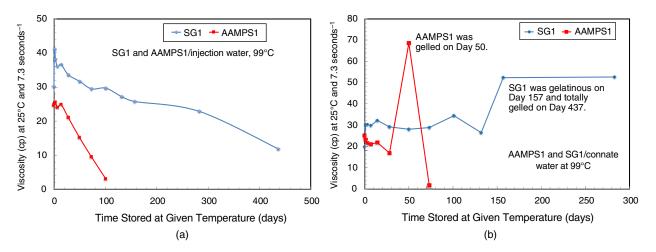


Fig. 8—Stability of AAMPS1 and SG1 at 99°C in (a) injection water and (b) connate water.

As mentioned previously, Davidson and Mentzer (1982), Kalpakci et al. (1990), and Jensen et al. (2018) reported good stability for scleroglucan in seawater at temperatures up to  $100^{\circ}$ C over a 1- to 2-year period, while Ryles (1988) noted that scleroglucan degraded within 3 months at 90°C. In view of these reported results, our stability experiments for scleroglucan were repeated. The replicate results basically confirmed that the polymer showed poor stability at 140°C and higher. Scleroglucan showed a half-life of approximately 20 days at 120°C.

Because of (a speculated) concern of a major change in the degradation mechanism between 100 and 120°C, the stability of SG1 was studied specifically at 99°C. The blue curve in Fig. 8a reveals that SG1 in injection water exhibited a viscosity half-life of approximately 1 year (consistent with our earlier Arrhenius projection). In the connate water (blue curve in Fig. 8b), the polymer became gelatinous at 157 days at 99°C and was totally gelled at 437 days. Previous experience indicated that gelatinous polymers will not flow effectively through porous media. If these results and those of the discussed previous researchers are taken at face value, it appears that scleroglucan might exhibit good stability at 99°C in seawater (3 to 4% TDS) but not in our more saline brines (6.9 to 24.4% TDS).

*Polymers with High ATBS Content.* Gaillard et al. (2015) provided generic compositions of polymers that contain acrylamide, acrylate, ATBS, and/or NVP. The polymers ATBSA, ATBSB, and ATBST contain very high ATBS levels. As of 621 days into the study, ATBSA exhibits excellent stability in all three brines at 120°C. In all three brines, the viscosity half-life was roughly 1 year at 140°C, 50 days at 160°C, and approximately 8 days at 180°C. Substantial gel was noted in the connate water after 84 days at 160°C. Achieving approximately 25 cp required 3100 mg/L in smartwater, 4100 mg/L in injection water, and 4200 mg/L in connate water.

Table 3 reveals that the polymers ATBSB and ATBST exhibit similar stability behavior as ATBSA. These three polymers all contain more than 90% ATBS. ATBSB has the same composition as ATBSA but a higher  $M_w$ , thus explaining why less polymer is needed to achieve the target viscosity (e.g., 2900 mg/L vs. 4100 mg/L in injection water). ATBST also has approximately the same composition as ATBSA, but it contains a small amount of a special monomer that reversibly promotes higher viscosities at high temperatures. All three polymers have more than 90% degree of anionicity because the ATBS monomer is negatively charged. Also, one should note that the ATBS monomer has approximately three times the  $M_w$  of acrylamide (i.e., 207 vs. 71 g/mol). Thus, for a given degree of polymerization, an ATBS polymer has approximately three times higher  $M_w$  as polyacrylamide. For ATBSB, achieving approximately 25 cp required 2700 mg/L in smartwater, 2900 mg/L in injection water, and 3100 mg/L in connate water. For ATBST, achieving approximately 25 cp required 2800 mg/L in smartwater, 3500 mg/L in injection water, and 3800 mg/L in connate water.

For the nine ATBS cases in Table 3, the projected half-lives at 99°C range from 55 to 133 years, with an average of 79 years and a standard deviation of 30 years. Note that all nine cases project a viscosity half-life exceeding 50 years at 99°C. For these same nine ATBS cases, the projected half-lives at 120°C averaged 5.6 years, with a standard deviation of 1.7 years. Within the uncertainty associated with the correlation coefficients in Table 4, it is unclear whether the three ATBS polymers have significantly different stabilities or that stability is different in the three brines. Stated another way, these three ATBS polymers exhibit excellent stability (i.e., the best ever reported for EOR polymers) at salinities ranging from 0.69 to 24.4% TDS and with divalent-cation levels from 0.0342 to 2.16% (from Tables 1 and 3). The study indicates that these three ATBS polymers should be sufficiently stable for potential applications with well spacings of 1 to 2 km, where polymer stabilities of 17 to 34 years would be desired. However, in an effort to promote the diversity of polymer vendors, only one of the three ATBS polymers was pursued during our coreflood studies.

For the very stable ATBSA polymers mentioned above, one could analyze the data in more pessimistic ways. For example, one could argue that a slow degradation mechanism is valid until the ATBS content is reduced (by hydrolysis) from near 100 to 80%, and then a faster degradation mechanism becomes important because close proximity to acrylate groups catalyzes hydrolysis of ATBS groups (Sandengen et al. 2018). With this assumption, one might disregard the stability data at 160 and 180°C and the data at 140°C after 150 days. Re-analyzing the data with these assumptions yields a more pessimistic projection for half-life at 99°C averaging 18.2 ( $\pm$ 6.5) years instead of >50 years from the original analysis (of data at all temperatures).

In Table 3, the polymers labeled ATBS8A and ATBS8B are thought to contain approximately 80% ATBS. For the six ATBS8 cases in Table 3, the projected half-lives at 99°C range from 0.4 to 4.5 years, with an average of 2.9 years and a standard deviation of 1.6 years. The ATBS8 polymers were expected to be less stable than ATBSA, ATBSB, and ATBST because they contain less ATBS (80% vs. more than 90%). Within the uncertainty associated with the correlation coefficients in Table 4, it is unclear whether either of the ATBS8 polymers has significantly different stabilities or that stability is different in the three brines. Visually from the data,

however, it appears that ATBS8A is more stable than ATBS8B, and the polymers are more stable in the less saline brine (i.e., smartwater). Although these polymers might not be sufficiently stable for the more demanding high-temperature/salinity applications with large well spacing, they could be sufficiently stable for other applications that have tighter well spacing.

AMNVPATBS1 contained 50 to 65% acrylamide, 20 to 25% ATBS, and 5 to 20% NVP. For the three AMNVPATBS1 cases in Table 3, the projected half-lives at 99°C were 1.3 years in smartwater, but only 0.1 year in injection or connate water. These findings are reasonably consistent with those reported by Gaillard et al. (2014) and Vermolen et al. (2011). It is doubtful that this polymer would be viable for the intended application. Nevertheless, this polymer was included during coreflood studies as a benchmark because it has been extensively studied internally, with some results reported by AlSofi et al. (2017a, 2017b). The decrease in stability from ATBS to ATBS8 to AMNVPATBS1 was expected because the ATBS content of the polymers decreased in that order.

*NVP Polymer (NVP1).* NVP1 is a powder-form polymer that contains a high level of NVP. NVP1 solutions have generally remained clear, except that the solution in the connate brine turned cloudy after 39 days at 180°C. In all three brines, the polymer solutions lost 40 to 60% of their original viscosity within 2 weeks at the tested temperatures. Also, the viscosity loss was greater as the storage temperature increased. We were surprised by this viscosity loss because NVP was expected to be a very stable monomer (Doe et al. 1987). After this initial viscosity loss, the decline in viscosity was much more gradual over the next 650 days at temperatures from 120 to 160°C. In examining the statistics for NVP1, the projected half-life of the polymer is 3.5 to 5 years in the three brines (Table 3). However, the correlation coefficients are marginal for most NVP1 cases (Table 4). Because of the influence of the rapid initial viscosity loss, this skewed the fits for this polymer to give, perhaps, an overly pessimistic view of the polymer's stability. At 99°C, the polymer might be considerably more stable than indicated by the projections. At present, the primary concerns with this polymer are that 7000 to 9100 mg/L of polymer are required to achieve the target viscosity, and the polymer is thought to be quite expensive.

Gaillard et al. (2014) offered an explanation for the sudden viscosity loss, followed by more gradual degradation. They pointed out that during polymerization of NVP copolymers, the early part of polymerization is dominanted by non-NVP monomers reacting first (producing polymers with low NVP content), while most of the NVP polymerization occurs later in the reaction. The implication is fraction of the product that contained little NVP (e.g., acrylamide) is susceptible to rapid degradation and viscosity loss, while the material with high NVP content experiences much slower degradation.

*Hydrophobic Associative Polymers (HYASA and HYASB).* Solutions of HYASA lost viscosity at moderate rates, depending on the temperature. It showed similarities to NVP1 in that a significant viscosity loss occurred within the first 2 weeks, followed by a more gradual viscosity decline. The projected viscosity half-lives at 99°C were 2.6 and 2.2 years in smartwater and injection water, respectively, but 5.4 years in connate water (Table 3). As with NVP1, the correlation coefficients (Table 4) were not as high as for other polymers. Therefore, the stability projections might be overly pessimistic. HYASB showed similar stability as HYASA in smartwater (3.3-year projected half-life at 99°C) but was notably less stable in the more saline brines. For HYASA, achieving approximately 25 cp required 6000 mg/L in smartwater, 8000 mg/L in injection water, and 6500 mg/L in connate water.

**Discussion of Coreflood Results.** *Retention.* Table 6 provides a summary of the coreflood results. In these experiments, the lowest polymer-retention value was 200  $\mu$ g/g. For a 4,100 ppm polymer solution, 200  $\mu$ g/g requires that the polymer bank be increased in size by 13% to achieve the same effect as when no polymer retention occurs (Manichand and Seright 2014). Of course, polymer concentrations and polymer-retention values as high as some of those observed in this work could jeopardize the viability of a polymer flood. Note that our work was conducted in carbonate cores with no residual oil saturation present. Wever et al. (2018) observed that polymer retention was 10 times lower at residual oil saturation than when no residual oil was present. In contrast, others (Wang et al. 2020) reported that retention was only modestly affected by the presence of residual oil. Future work will be performed to examine whether more palatable polymer-retention values will occur under our conditions when residual oil is present. We acknowledge that the presence of residual oil might have an accentuated effect on the retention of hydrophobic associative polymers.

**Residual Resistance Factors and Plugging.** Relatively short cores were available for this work (Table 5), so no internal pressure taps were used. Thus, if any face plugging occurred (whether immediate or prolonged), we could not distinguish face plugging from real resistance factors. Fortunately, for all but one of our floods, no continuous or sustained plugging occurred. Also, in three of the cases, residual resistance factors were low (approximately 2), suggesting little face plugging (last row of Table 6). For two cases in Table 6, residual resistance factors ranged from 4 to 6.5, suggesting either that some face plugging occurred or that some internal plugging of the core occurred. In these cases, we did not observe progressive plugging during polymer injection. The SG1 scleroglucan case was the only flood where severe progressive plugging was noted during polymer injection.

*ATBSA*. In 145-md carbonate rock, ATBSA exhibited polymer retention of 211  $\mu$ g/g, no plugging while injection, virtually no thermal degradation during injection at 1 to 2 ft/D, and a residual resistance factor of 2.2. In 24-md carbonate, ATBSA exhibited polymer retention of 415  $\mu$ g/g, no plugging during injection, 7% viscosity loss during injection at 1 to 2 ft/D, and a residual resistance factor of 6.5. As mentioned previously, at 99°C, the projected viscosity half-life was more than 50 years for this polymer.

Masalmeh et al. (2019) evaluated ATBSA at 105 and 130°C in carbonate cores. Consistent with our results, they found that the polymer showed good stability over 6 months at 130°C and over 1 year at 120°C. As in our experiments (Fig. 6), they also noted shear thickening in porous media and good resistance to mechanical degradation. They reported polymer retention of 115  $\mu$ g/g in a 263-md carbonate core that was aged in the presence of crude oil. In cores with no oil, they found retention values of 316  $\mu$ g/g in 121-md carbonate, 209  $\mu$ g/g in 45-md carbonate, and 570  $\mu$ g/g in 29-md carbonate. These values are reasonably consistent with our value of 211  $\mu$ g/g (measured with no residual oil saturation) in 153-md carbonate.

Alfazazi et al. (2019) also examined ATBSA at 25 and 90°C in Indiana limestone cores. Most of their results were quite consistent with ours. Their retention values ranged from 194 to 340  $\mu$ g/g in 143- to 153-md carbonate, which brackets our value. They did report two differences in behavior. First, they reported no shear thickening for ATBSA at 90°C. Second, they noted a substantial degree of mechanical degradation. They suggested that the degradation might be attributable to passing through a backpressure regulator.

Our results in 24-md carbonate are also reasonably consistent with those reported by Masalmeh et al. (2019). In particular, our retention value of 415  $\mu$ g/g in 24-md carbonate is consistent with their value of 570  $\mu$ g/g in 29-md carbonate. In 44-md carbonate, Hashmet et al. (2017) noted a residual resistance factor of 1.73 associated with a 0.1-PV polymer bank.

*AMNVPATBS1*. AMNVPATBS1 exhibited polymer retention of 911 µg/g, no plugging, a 9% viscosity loss during injection at 1 to 2 ft/D, and a residual resistance factor of 2.2. The projected viscosity half-lives at 99°C were 1.3 years in smartwater and 0.1 year in injection water. In contrast to our polymer retention of 911 µg/g, Gaillard et al. (2014) reported retention of 84 µg/g for a similar polymer in carbonate cores under somewhat different conditions. For comparison, AlSofi et al. (2017b) reported HPAM retention values from 155 to 530 µg/g in carbonate cores at 99°C. Our AMNVPATBS1 residual resistance factor was 2.2. Gaillard et al. (2014) reported

residual resistance factors for a similar polymer in the range of 1.1 to 1.3 in carbonate cores under somewhat different conditions. For comparison, HPAM and sulfonated polyacrylamide (containing approximately 25% ATBS) in seawater exhibited low residual resistance factors in carbonate cores at 99°C (Han et al. 2012; AlSofi et al. 2017b).

*SG1 Scleroglucan.* SG1 scleroglucan showed no mechanical degradation or shear-thickening behavior at 99°C. However, it plugged the core during injection, exhibited polymer retention exceeding 1250  $\mu$ g/g, and experienced a 70% viscosity loss while transiting the core at 1 to 2 ft/D.

Rivenq et al. (1992) reported scleroglucan retention of 30  $\mu$ g/g at 90°C in a Berea Sandstone core (in seawater). In NaCl solution, Audibert et al. (1993) also reported scleroglucan retention of 30  $\mu$ g/g in Berea Sandstone at 90°C. Fournier et al. (2018) reported retention of 8  $\mu$ g/g in an Estaillade limestone core at 50°C. In contrast, Huang and Sorbie (1993) reported retention values more than 80  $\mu$ g/g (accompanied by apparent face plugging) in sandpacks at room temperature. Kulawardana et al. (2012) found that scleroglucan propagated through Berea Sandstone more rapidly (i.e., earlier arrival of the polymer front) as temperature increased from 25 to 100°C. Although they did not report polymer-retention values, examination of their data reveals that scleroglucan retention was quite high at 100°C. For schizophyllan (a polymer that is structurally similar to scleroglucan), Quadri et al. (2015a, 2015b) reported low retention values (0.53 to 47.5  $\mu$ g/g) in 3- to 165-md carbonate cores at 120°C.

If the literature reports and our data are taken at face value, it appears that scleroglucan can show very good performance at high temperatures under some conditions of salinity (e.g., seawater), hardness, and core material, but not others. Under our particular conditions (6.9% TDS, 0.34% divalent cations, carbonate core), scleroglucan did not propagate well.

*NVP1*. NVP1 showed polymer retention of 204  $\mu$ g/g, no mechanical degradation, little or no plugging during injection, and a residual resistance factor of 4. However, the polymer experienced 23% viscosity loss during injection at 1 to 2 ft/D. This polymer required the highest concentration (9000 mg/L) to achieve the target viscosity. The projected half-life at 99°C was 3.7 years.

Doe et al. (1987) reported retention for this type of polymer in seawater at  $121^{\circ}$ C to be approximately 250  $\mu$ g/g in approximately 100-md Berea sandstone and approximately 100  $\mu$ g/g in approximately 100-md sandstone cores from the North Burbank unit (all at residual oil saturation).

*HYASA*. For HYASA at 99°C, the projected viscosity half-life ranged from 2.2 to 5.4 years, depending on the brine. This polymer exhibited a retention of 1126  $\mu$ g/g, but no significant viscosity loss or plugging during flow through a 504-md carbonate core at 99°C. The residual resistance factor was 2.3. Reichenbach-Klinke et al. (2011) speculated that low residual resistance factors for polymers might indicate low polymer retention. However, results here and elsewhere (Wang et al. 2020) provide counterexamples to this idea.

## Conclusions

An extensive evaluation was performed to identify polymers for use in the polymer flooding of high-temperature carbonate reservoirs with hard, saline brines. These polymers included new ATBS polymers, scleroglucan, NVP-based polymers, and hydrophobic associative polymers. We present the following conclusions:

- 1. Three ATBS polymers were identified that were projected to provide a viscosity half-life over 50 years at 99°C (under oxygen-free conditions) in the hard, saline brines of interest for the potential project. Other polymers examined in this study could not meet the required minimum of a 17-year half-life at 99°C.
- 2. During corefloods using carbonate cores under oxygen-free conditions at 99°C, polymer-retention values were relatively high for all polymers examined, with the lowest values being approximately 200  $\mu$ g/g.
- 3. During the corefloods, only scleroglucan exhibited significant plugging during injection of polymer solutions at 99°C.
- 4. For all coreflood cases where plugging was not observed, resistance factors were consistent with expectations from viscosity measurements.
- 5. Three of five polymers showed modest or no viscosity loss in passing through cores at low velocity at 99°C. As exceptions, one polymer (scleroglucan) showed substantial viscosity loss because of polymer being stripped from the solution, while a synthetic NVP-based polymer experienced significant thermal degradation.
- 6. Residual resistance factors were moderate for cases where severe face plugging was not observed.

## Nomenclature

- $A_1$  = pre-exponential factor in Eq. 1, unitless
- $A_2$  = pre-exponential factor in Eq. 2, unitless
- C = polymer concentration, ppm
- E =activation energy, cal/mol
- $k_w$  = absolute permeability to water, darcy [ $\mu$ m<sup>2</sup>]
- $M_w$  = polymer molecular weight, g/mol [dalton]
- R = universal gas constant, cal/mol
- t = time, days
- T =temperature, K
- $\mu = \text{viscosity}, \text{cp} [\text{mPa} \cdot \text{s}]$
- $\mu_o$  = viscosity at time zero, cp [mPa·s]
- $\mu_s =$ solvent viscosity, cp [mPa·s]
- $\tau = \text{decay constant, days}$
- $\tau_{99^{\circ}C} = \text{decay constant at } 99^{\circ}\text{C}, \text{days}$ 
  - $\phi = \text{porosity}$

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SI Metric Conver	sion Factors
$cp \times 1.0^*$	$E - 03 = Pa \cdot s$
$ft \times 3.048*$	E - 01 = m
in. $\times 2.54*$	E+00 = cm
$md \times 9.869\ 233$	$E-04 = \mu m^2$
psi × 6.894 757	E+00 = kPa
*Conversion is exact.	